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(54) Title: NON-MAMMALIAN GnRH ANALOGS AND USES THEREOF IN TUMOR CELL GROWTH REGULATION AND CANCER THERAPY

(57) Abstract: Specially designed non-mammalian GnRH analogs resistant to degradation by the tumor tissue enzymes, post-proline peptidases as well as endopeptidases, are disclosed. The GnRH analogs are further defined as analogs of chicken II GnRH, salmon GnRH, or herring GnRH, but can include any non-mammalian GnRH analog with similar amino acid structure. These non-mammalian analogs incorporate D-arginine, D-leucine- D-t-Bu-Serine or D-Trp or other similar amino acids at position 6 and ethylamide or aza-Gly-amine or similar amides at position 10. These analogs demonstrate preferential binding to tumor cell GnRH receptors that is greater relative to the binding of the mammalian analogs to the tumor cell GnRH receptor. These non-mammalian GnRH analogs may be used in pharmaceutical preparations, and specifically in various treatments as an anti-tumor, anti-proliferation, anti-metastatic and/or apoptotic agent. The non-mammalian GnRH analogs are also provided in pharmaceutical preparations that may be used clinically for tumor regression when used in very low doses and administered in pulsatile fashion.

Title: Non-Mammalian GnRH Analogs and Uses Thereof in Tumor Cell Growth Regulation and Cancer Therapy

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BACKGROUND OF THE INVENTION

1. Field of the Invention

Applicants' invention relates to the field of tumor growth regulation. More particularly, Applicants' invention concerns unique non-mammalian peptide hormone analogs of non-mammalian gonadotropin releasing hormone (GnRH) and the method for use of these analogs in the regulation of cell growth, particularly cancer cell growth.

2. Background Information

Gonadotropin-releasing hormone (GnRH) is a hormone known to be produced in the hypothalamus with binding affinity for the pituitary gland. When hypothalamic GnRH binds to the pituitary it causes the pituitary gland to release the gonadotropins (i.e. gonad stimulating hormones) luteinizing hormone (LH) and follicle-stimulating hormone (FSH). Each of these pituitary hormones has a different effect depending on one's sex. One important effect is the production and secretion of the gonadal steroids estrogen and progesterone in sexually mature females and testosterone in males.

Before the chemical characterization of the mammalian GnRH it was realized that a hypothalamic substance regulated the production of pituitary LH and FSH (1) and that these gonadotropins regulated gonadal steroidogenesis. The delineation of mammalian GnRH enabled Applicants to synthesize this decapeptide and administer it systemically to humans (2). It was then recognized that a long acting superagonist of this mammalian GnRH effected a flare release of pituitary gonadotropins followed by their inhibition (3). The inhibition was effected by a down-regulation of the pituitary GnRH receptor which corresponded downstream to an inhibition of the gonadal steroids, estrogen, progesterone and/or testosterone.

This is a form of chemical castration.

Since certain types of tumors, such as certain breast (4) and prostate cancers (5), are now known to be dependent on gonadal steroids, mammalian GnRH analogs have been used to suppress gonadal steroids via their chemical castration activity (3). Thus, we know that the use of mammalian GnRH analogs is feasible as a treatment of certain cancers.

It was only with the development of sensitive and specific radioimmunoassays for GnRH and GnRH-like molecules that a very surprising finding was reported. That finding was initially described by Applicants. Applicants reported that GnRH-like molecules exist and function not only in the hypothalamic-pituitary axis, which functions as an endocrine system to distribute hormone systemically, but GnRH-like molecules also exist in extra-hypothalamic tissues(6-9) to provide a paracrine action i.e. localized signal secretion. It is now realized that paracrine action of GnRH-like substances have functions in the placenta, gonad, breast, prostate, etc., (10-13) as well as in many cancerous tumors (14-31). Even with this general knowledge, the effective use of mammalian GnRH analogs to act directly on particularly tumor tissue has not resulted. One of the goals of the present invention was to utilize novel forms of GnRH not previously envisioned for cancer therapy that bind to the tumor GnRH-like receptor with 50 to 1000 fold the activity of mammalian GnRH or its superagonist and have potent bioactivity in inhibiting tumor cell growth.

The initial studies on GnRH activity in tumor tissues and the human placenta utilized mammalian GnRH and its analogs, in accordance with the teaching that the human encodes for only one isoform of GnRH (32,33). In Applicants' studies in the human placenta Applicants localized and quantified the concentration of GnRH produced by the human placenta throughout pregnancy (34,35). Applicants demonstrated the synthesis and activity of a GnRH-like molecule in regulating human chorionic gonadotropin (hCG) and steroid production (36-38), and the release of a GnRH-like molecule into the maternal circulation (34). Applicants also demonstrated that high doses of mammalian GnRH could stimulate hCG and

prostanoid production in a specific receptor mediated event and that GnRH-like production and activity in the human placenta is regulated by feedback interactions of estrogens and progesterone production (39-41). Thus, Applicants described and established the first paracrine system in human or mammalian physiology (42-45).

5 Concomitantly, Dubois et al. (12) described a second paracrine system from the presence of somatostatin in the pancreas. Thereafter, Applicants and other investigators reported actions of mammalian GnRH on placental function and identified feedback interactions including activin, inhibitin, follistatin, neurotransmitters, prostaglandins, and steroids (46-63).

10 Using in situ localization a message to mammalian GnRH has been localized at the syncytio- and cytotrophoblast, as well as the stroma of the placenta (64-66). A gene for mammalian GnRH differing from hypothalamic GnRH by the inclusion of the first intron and a very long first exon has been reported (67-69). Multiple transcription sites have been identified for the GnRH gene in the placenta as well
15 as in other reproductive tissues (70-72). Steroid regulatory sites on the promoter have also been identified (73,74). The functionality of the promoter is supported by the demonstration that mRNA for GnRH can be regulated by steroids (75-78).

Placental GnRH receptor activity and a GnRH mRNA have also been identified (79,80). The receptor number is highest in early gestation and down-
20 regulated by 12-20 weeks, and still detectable in term placenta, although the mRNA for mammalian GnRH is not (79,80). This pattern of receptor activity parallels that of chorionic GnRH-like activity (6,34) and supports the hypothesis that chorionic GnRH may down-regulate its receptor, as does mammalian GnRH and its analogs at the pituitary level. Studies of Szilagy et al. (81) and Currie et al. (82)
25 have indicated down-regulation of chorionic receptors by mammalian GnRH analogs. In addition, estradiol has been shown to upregulate the placental GnRH receptor. Thus, there is substantial data to indicate a functional, regulated GnRH receptor in extrahypothalamic tissues.

In total, these studies have firmly established the presence of a
30 hypothalamic-pituitary-gonadal axis in extra-hypothalamic tissue. Presently many

other hypothalamic-like activities, such as by corticotropin-releasing hormone (CRH), have now been defined in the placenta and other tissues as well. Such paracrine axis are known in the pancreas, thyroid, gut, bone, brain, ovary, endometrium, eye, etc.

5 Of particular interest to this invention are previous reports of the presence of GnRH-like substances and receptors in numerous cancer tissues and their cell lines (15,17,20,23,25,26,30,31,83,84). GnRH-like activity and its receptors have been identified in the breast, bronchial, ovarian, endometrial, prostate, gastrointestinal tumors. The function of a GnRH-like substance and its receptors in tumor tissues
10 is supported by the demonstration that mammalian GnRH can stimulate hCG from human and animal tumors and can inhibit cell growth in vitro. These findings have led to numerous studies of the effects of mammalian GnRH analogs on the expression of GnRH receptors, cell signal transduction, apoptosis, and overall growth of tumor cell lines (14,16,18,19,21,22,29,85-89). The growth of tumors
15 in vivo has also been studied with individual case reports of patients responsive to mammalian GnRH analogs (24,27,28,90-92).

However, some very problematic findings from previous studies in both the placenta and tumor tissue has led to skepticism about the true role of mammalian GnRH analogs in both tissues. The GnRH receptor affinity for GnRH in both the
20 placenta and tumors is on the order of 10^{-5} to 10^{-6} M (84,93,94). The biological significance of such a weak affinity in light of much lower levels of endogenous GnRH-like activity must be questioned. In addition, Applicants have observed in human pregnancy studies, both in vitro and in vivo, that mammalian GnRH appears to act as a partial agonist not a true agonist of tumor GnRH(38,95). When
25 receptors are available, mammalian GnRH acts as an agonist of tumor GnRH, but when tumor receptors are low or occupied, mammalian GnRH competes with the more potent tumor GnRH resulting in a partial agonist action. Furthermore, Applicants and others have observed that certain antibodies for mammalian GnRH reacted with chorionic GnRH with a different affinity (96-99). These findings led
30 Applicants to propose that neither the extra-hypothalamic GnRH nor its receptor are

identical to mammalian GnRH and its pituitary receptor (100,101).

Applicants have defined yet another difference in extra-hypothalamic GnRH, i.e., its metabolism. The metabolism of a hormone is as important for maintaining biologically active concentrations of that hormone, as that which stimulates the hormone's synthesis and release. For GnRH, in the non-pregnant human, both in the pituitary and in the circulation, the predominant enzymatic degradation is directed to the 5-6 peptide bond catalyzed by an endopeptidase. Thus, existing analogs of the mammalian GnRH each bear a D-amino acid substituted in the 6 position. However, Applicants have isolated and characterized the dominant enzyme that degrades GnRH in the placenta and it is a post-proline peptidase acting to cleave the proline-glycine peptide bond at the 9-10 position (102,103). Applicants have recently obtained similar data for the metabolism of GnRH in breast tumor cells. Thus, there appears to be cell specific metabolism of GnRH at the placenta and breast tumor cells which differs from that in blood and the pituitary.

Since it appeared as though there was a different form of GnRH at work at the placenta and breast tumor cells, various isoforms of GnRH were investigated. Different isoforms of GnRH have been identified in non-mammalian species, such as fish and aves. The unique sequence of these GnRH are known. Chicken I, chicken II, salmon, catfish, dogfish, lamprey and more recently herring GnRH have also been reported (33,106). In lower vertebrates a number of GnRH isoforms can be expressed in the same species (32,33,76,78,105,107-116). In most cases, each decapeptide conserves the first four, the sixth and in every case, the last two amino acids in the GnRH molecule, but have varying amino acids in the fifth, seventh and/or eighth position. These modifications render the molecule unique, having only weak affinity for the mammalian pituitary receptor, although conversely mammalian GnRH is active in many lower vertebrate classes.

As mentioned, in certain lower vertebrates a number of GnRH isoforms are expressed in the same species. In amphibians, a chicken II GnRH receptor as well as a mammalian GnRH receptor has been reported. However, it was not until 1994, when Dellovade et al. (117) and King et al. (118) described chicken II GnRH in

musk shew, mole and bat brain, that the existence of multiple isoforms of GnRH in a mammal was realized. Even then, it was still thought that modern placental mammalian species did not encode or express other than mammalian GnRH. Recently however, chicken II GnRH has been characterized in the tree shew (119),
5 guinea pig, and primate brain (120) and their separate genes have been described (121,122). Only this year has the code for the chicken II GnRH receptor been identified in human tissues.

In contrast, Applicants have proposed and obtained substantial data to support the hypothesis that non-mammalian isoforms of GnRH and their specific
10 receptors are expressed in extra-hypothalamic tissues and that the non-mammalian GnRH molecules are the true ligands for these receptors (123,124). Applicants have also proposed that these GnRH molecules have specific roles in regulating cell growth and cell death and are pivotal in regulating cell growth of GnRH responsive tumors by a direct receptor mediated action on these tumor cells.

15 It is believed that the non-mammalian GnRH isoforms and analogs of the present invention may act either as a superagonist at the tumor tissue leading to tissue receptor down-regulation, or as a pure antagonist of the endogenous isoform of GnRH in the tumor tissue, acting via the tumor tissue receptor. The down-regulation of the GnRH receptor or the antagonism of the endogenous isoform of
20 GnRH will provide for a reduction in cell proliferation and/or induce apoptosis. The specific action of the non-mammalian GnRH analog will compete at the tumor cell GnRH receptor(s) with the endogenous isoform of GnRH effecting an antagonism or a superagonistic down-regulation of the receptor, leading to cell death and regression of the tumor and inhibition of metastasis. Thus, this agent may be used
25 to reduce tumor growth. To date, no such non-mammalian GnRH analog has been designed which has stability and tumor tissue specificity.

To date, little if any data, has been reported in relation to non-mammalian GnRH activity on tumor tissues. Chicken I GnRH and Lamprey GnRH (18,86,125,126) have been studied and limited activity was found. Applicants
30 have studied these isoforms of GnRH and have found no or limited binding activity

in chorionic tissues. On the other hand, Applicants have demonstrated greatly enhanced binding and bioactivity of chicken II GnRH and salmon GnRH analogs as compared to mammalian GnRH or its analogs in both breast cancer cells and placental tissue. Thus, Applicants have obtained data to support the hypothesis
5 that certain non-mammalian GnRH analogs have enhanced receptor and bioactivity for tumor tissues and this finding taken together with the understanding of the unique metabolism of GnRH isoforms in cell specific sites have formed the basis of Applicants invention, i.e., the utilization of stable, cell-active analogs of non-mammalian GnRH isoforms to regulate tumor cell growth and the treatment of
10 cancer. In addition, Applicants postulate that due to similar amino acid structures, Herring GnRH, Dogfish GnRH, and Catfish GnRH as well as other GnRH isoforms and analogs with similar amino acid structure should exhibit the same or similar binding and bioactivity.

SUMMARY OF THE INVENTION

15 It is an object of the present invention to provide a novel pharmaceutical preparation that includes non-mammalian GnRH isoforms and their analogs specifically designed to bind to extra-pituitary GnRH receptors expressed on tumor tissues.

It is another object of the present invention to provide novel GnRH analogs
20 that are resistant to degradation by post-proline peptidases, particularly those which are known to be found around tumor tissues.

Still another object of the present invention is to provide novel GnRH analogs that are resistant to the endopeptidase found circulating in the blood.

Another object of the present invention is to provide novel GnRH analogs
25 that act as superagonists at the tumor tissue leading to tissue receptor down regulation.

Yet another object of the present invention is to provide novel GnRH analogs that act as pure antagonists of the endogenous form of GnRH in tumor tissue via the tumor tissue receptor.

30 Another object of the present invention is to provide novel GnRH analogs

which reduce tumor cell proliferation.

Still another object of the present invention is to provide novel GnRH analogs which induce apoptosis.

It is yet another object of the present invention to provide novel GnRH
5 analogs which reduce tumor cell metastasis.

Another object of the present invention is to provide novel GnRH analogs that can be used as anti-tumor agents.

Yet another object of the present invention is to provide novel GnRH analogs that can be used to reduce tumor cell growth.

10 It is still another object of the present invention to provide novel GnRH analogs that are stable in circulation.

Another object of the present invention is to provide novel GnRH analogs that are tumor tissue specific.

Yet another object of the present invention is to provide novel GnRH analogs
15 which can be used to induce tumor regression.

Still another object of the present invention is to provide a novel method for synthesizing analogs of non-mammalian GnRH isoforms having increased activity in tumor tissues.

It is another object of the present invention to provide a novel method for
20 inhibiting tumor growth which in turn reduces tumor cell proliferation, tumor size and metastasis i.e. apoptosis and tumor regression.

It is yet another object of the present invention to provide a novel method for using non-mammalian GnRH analogs directly on tumors as an anti-tumor or anti-metastasis drug.

25 Another object of the present invention is to provide a novel non-mammalian GnRH analog composed of Salmon, Chicken II, or Herring GnRH analogs that are modified at the C-terminal and which shown greater affinity for the tumor receptor than mammalian GnRH.

It is still another object of the present invention to provide a novel non-
30 mammalian GnRH analog which has an aza-Gly-NH₂ substitution at the number 10

position to make the sequence more stable in tumor tissues and in blood and to inhibit degradation by post-proline peptidases.

Yet another object of the present invention is to provide a novel non-mammalian GnRH analog sequence which is substituted at the 6 position with preferably a D-Arg but could be any other D-amino acid such as, but not limited to, D-Leu, D-Trp, and D-Bu-Ser.

It is another object of the present invention to provide a novel non-mammalian GnRH analog which has increased binding affinity to the tumor receptor and metabolic stability.

Still another object of the present invention is to provide a novel non-mammalian GnRH analog which will not be toxic after long term therapies.

Yet another object of the present invention is to provide a novel more potent non-mammalian GnRH analog which can be used to bind to the tumor tissue GnRH receptor with high affinity so as to displace the endogenous GnRH-like activity and block its action.

It is another object of the present invention to provide a novel non-mammalian GnRH analog which incorporates a substitution of Gly(10)-NH₂ with ethylamide to inhibit degradation by post-proline peptidases.

Still another object of the present invention is to provide a novel non-mammalian GnRH analog which has minimal effect on the mammalian pituitary GnRH receptor.

In satisfaction of these and related objectives, Applicant's present invention provides unique non-mammalian peptide hormone analogs of non-mammalian GnRH and the method for use of these analogs in the regulation of cell growth, particularly tumor cell growth. Applicant's invention permits its practitioner to treat patients who have cancerous tumors with non-mammalian GnRH analogs which have high binding affinity to the GnRH receptors located on tumor cells which in turn reduces the tumor size and proliferation.

BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1. Chart of the sequences of known isoforms of GnRH.

Figure 2. Effect of des-Gly(10)-mammalian GnRH-ethylamide on the degradation of mammalian GnRH by the chorionic post-proline peptidase.

5 ◇ GnRH 0.00313 M, ▽ GNRH 0.0625 M, ● GNRH 0.0125 M.

Figure 3. Action of D-Arg- Chicken II-aza-Gly-NH₂ on the Degradation of Mammalian GnRH by Chorionic Post-Proline Peptidase. ◇ GnRH 0.00313 M, ▽ GNRH 0.0625 M, ● GNRH 0.0125 M, ○ GNRH 0.0250 M.

Figure 4. Binding of I¹²⁵-D-Arg(6)-Chicken II GnRH-aza-Gly(10)-amide to MCF 7
10 breast cancer cells after 24 hours of incubation with no exogenous GnRH or competing isoforms or analogs of GnRH

Figure 5. The anti proliferative, tumor regression activity of D-Arg(6)-Chicken II GnRH-aza-Gly(10)-amide compared to controls and other isoforms and analogs of GnRH after 24 hours.

15 **Figure 6.** Inhibitor constants for analogs of GnRH.

DETAILED DESCRIPTION OF THE PREFERRED EMBODIMENT

Following long-standing patent law convention, the terms "a" and "an" mean "one or more" when used in this application, including the claims.

It should be appreciated by those of skilled in the art that the techniques
20 disclosed in the material which follows represent techniques discovered by the Applicants to function well in the practice of the invention, and thus can be considered to constitute preferred modes for its practice. However, those skilled in the art should, in light of the present disclosure, appreciate that many changes can be made in the specific embodiments which are disclosed and still obtain a like or
25 similar result without departing from the spirit and scope of the invention.

Design & Synthesis of Non-Mammalian GnRH Analogs

The preferred embodiment for the present invention is the design of non-mammalian GnRH analogs that have increased activity in tumor tissue cells by exhibiting potent tumor receptor binding activity, tumor tissue stability, and
30 biological activity.

Existing mammalian GnRH analogs are designed for activity at the pituitary GnRH receptor and with extended stability in the circulation of non-pregnant individuals i.e. protection from endopeptidase degradation. Yet, existing data indicate that the tumor GnRH receptor differs from that in the pituitary. Therefore, prior known pituitary mammalian GnRH analogs have not been designed for direct use in tumor tissues and potent non-mammalian GnRH analogs have not been designed for use to regulate cancer cell growth. The present invention provides potent non-mammalian GnRH analogs that act directly on tumor cell growth and proliferation.

Non-mammalian analogs of GnRH were designed according to general guidelines established by Applicants. First, these analogs were specifically designed to be resistant to degradation both in the maternal circulation as well as within the tumor tissue by endopeptidases and post-proline peptidases. This allows for the maintenance of sufficient concentrations of analog when administered via the maternal system to reach the cancerous tissue. And second, the analogs were designed according to the particular specificity of the tumor cell receptor to specifically bind the GnRH receptors with high affinity so as to preferably displace the endogenous GnRH activity and block its action. This tumor GnRH binding specificity can effect either a down regulation of the tumor GnRH receptor or act as a true antagonist to inhibit tumor growth, proliferation, and metastasis by inducing apoptosis and tumor regression by directly inhibiting tumor function.

Analogues of the Salmon and Chicken II GnRH sequences, that show greater affinity for the tumor GnRH receptor than for the pituitary GnRH receptor, were modified to the aza-Gly(10)-NH₂ analog to make them resistant to degradation by post-proline peptidases (See Figure 1, Analogs 2, 4 and 6). Ethylamide or other similar amides can be used at position 10. Next, the Chicken II GnRH sequence, and the Salmon GnRH sequence were modified at the 6 position using D-Arg, D-Leu, D-tBu-Ser, D-Trp, or other similar amino acid making the analog resistant to degradation by the endopeptidase in blood, and at the 10 position using preferably aza-Gly(10)-NH₂, making it stable in maternal blood and the tumor tissues (See

Figure 1, Analogs 2, 4 and 6). Again, ethylamide or other similar amides can be used. These analogs are expected to have not only increased binding to the tumor receptor but also increased metabolic stability. Due to the similarity in amino acid makeup, these procedures could be repeated using Herring GnRH, Dogfish GnRH, and Catfish GnRH as well as any other decapeptide with similar amino acid structure. The preparation and chemical manipulation of these non-mammalian GnRH analogs can be completed with any standard protocol.

Tumor Receptor Binding of GnRH Isoforms and Analogs

The tumor receptor binding activity of the different non-mammalian GnRH analogs of the present invention was compared. Prior mammalian GnRH analogs have been designed to increase activity at the pituitary GnRH receptor and stability in the circulation of nonpregnant individuals. These analogs do not demonstrate as potent binding activity at the tumor receptor as they do at the pituitary receptor. In contrast and as was mentioned earlier, the non-mammalian GnRH analogs of the present invention have been designed to interact with preference at the tumor receptor and not the pituitary receptor.

GnRH receptors on the cells of MCF-7 breast cancer cells were studied. The cells were plated on 96 wells and grown to confluency in base medium (M3D:Fetal Bovine Serum [9:1]). Prior to the experiment the cells were down shifted to M3D:Fetal Bovine Serum [99:1] and then to serum-free medium. Cells were incubated for 24 hours at room temperature with mammalian GnRH, Buserilin, Leuprolide, Chicken II GnRH, and D-Arg(6) Chicken II-aza-Gly(10)-amide. Cells were then collected and studied for receptor binding and receptor number with D-Arg(6) Chicken II-aza-Gly(10)-amide. Addition of enzyme inhibitors of the endogenous post-proline peptidase and other peptidases were used as well as agents for receptor stabilization. Receptor bound label was separated by centrifugation. The binding of the non-mammalian GnRH analog and its ability to regulate the tumor cells' GnRH receptor was compared. Figure 4 shows the binding of 125 I-D-Arg(6)-Chicken II GnRH-aza-Gly (10) -amide to MCF-7 breast cancer cells after 24 hours of incubation with no exogenous GnRH or competing

isoforms or analogs of GnRH. D-Arg(6) -Chicken II GnRH-aza-Gly(10)-amide specifically bound the MCF-7 breast cancer cells. This binding was competitively inhibited by D-Arg(6)-Chicken II GnRH-aza-Gly(10)-amide with the greatest potency. This was followed by Buserilin. Mammalian GnRH was the weakest competitor, while Chicken II GnRH was highly potent even though it is not protected from degradation either at the 6 or the 10 position. This indicated the high innate affinity of this isoform of GnRH for the tumor GnRH receptor. It is believed by Applicants that the same or similar results could be obtained using other non-mammalian GnRH isomers or analogs with similar amino acid structure as Chicken II GnRH such as but not limited to Salmon GnRH, Herring GnRH, Catfish GnRH, or Dogfish GnRH.

Tumor Tissue Stability Studies for GnRH Isoforms and Analogs

As has been previously mentioned, mammalian GnRH and its analogs bind with weak affinity to tumor cell receptors in certain tumor tissues whereas the non-mammalian GnRH analogs exhibit strong affinity for these receptors. Observing this strong affinity it became necessary to examine the non-mammalian GnRH analogs for stability. The non-mammalian GnRH analogs of the present invention have not previously been examined for stability. However, the added stability of these non-mammalian analogs would effect a substantial increase in bioactivity. Thus, stability studies involving endopeptidase and post-proline peptidase were performed for the non-mammalian GnRH analogs.

Endopeptidase Stability Studies:

Since human pituitary and blood contain an enzymatic activity that degrades GnRH at the 5-6 position, rather than the 9 position, present non-mammalian GnRH analogs have been designed to inhibit the former enzymatic activities and have substitutions in the 5-6 position of the molecule. The present analogs with these substitutions are therefore resistant to degradation at the pituitary or in the blood of non-pregnant individuals. However, these substitutions alone do not protect the analogs from degradation at the tumor tissues which contain post-proline peptidase. Substitution of the Gly(10)-NH₂ with ethylamide, or the more potent

aza-Gly(10)-NH₂, inhibits degradation by post-proline peptidases. A number of existing mammalian GnRH analogs also have a substitution of Gly(10)-NH₂.

Post-Proline Peptidase Stability Studies:

As mentioned earlier, the post-proline peptidase is important in actively
5 degrading peptides that contain a proline residue. GnRH is such a peptide. Initially the enzymatic activity of the tumor cell was studied. Tumor tissue cells and their spent media were studied for enzyme activity. In particular, examination was made for the degradation of GnRH both with and without specific post-proline and endopeptidase activity inhibitors to determine the specificity of the tumor enzymatic
10 activity. These studies have demonstrated very high post-proline peptidase activity produced by the tumor tissue.

The enzymatic degradation of the non-mammalian GnRH analogs were studied in MCF-7 breast cancer cells using an enzymatic activity assay and compared to that for the purified chorionic post-proline peptidase. Chorionic post-proline
15 peptidase is a peptidase with high specificity for the degradation of GnRH at the proline-glycine bond, but can also degrade other GnRH species containing this bond.

In a non-pregnant individual very little post-proline peptidase activity is present in the blood or the pituitary. Thus, currently available mammalian GnRH
20 analogs have not been designed to be resistant to degradation by this activity. However, due to the high post-proline peptidase activity present in tumor tissue, the non-mammalian GnRH analogs for cancer therapy described herein were designed to be resistant to this type of degradation. The stability of these non-mammalian GnRH analogs in the presence of post-proline homogenates was examined and
25 compared to existing mammalian GnRH analogs. In addition, the ability of the analogs to competitively inhibit the degradation of GnRH using chorionic post-proline peptidase was studied.

The stability of most potent receptor-active non-mammalian GnRH analogs in the presence of tumor tissue cells, spent media, or tumor tissue cells homogenates
30 was identified. Using the incubation system developed for chorionic post-proline

peptidase activity, the degradation of GnRH was tested. Each of these analogs were first studied for their ability to act as a competitive inhibitor of GnRH for chorionic post-proline peptidase activity using the enzymatic activity assay as described previously (103). In this assay, incubation of enzyme and mammalian GnRH with
5 and without the chosen newly synthesized non-mammalian GnRH analog was studied. The reaction was stopped by heating at 85°C for 10 minutes. The remaining mammalian GnRH substrate was quantified by radioimmunoassay. The product formed, i.e. the N-terminal nonapeptide of GnRH, was calculated by subtraction, and its inverse plotted against the inverse of the original substrate
10 concentrations to determine the Ks of the competition. The Ki was determined by plotting the inverse of the product that formed versus the inhibitor used. The inhibitory activity of Antide, Im-btl-D-His(6)-mammalian-GnRH-ethylamide, D-Trp(6)-GnRH-ethylamide, Buserilin, Leuprolide, OH-Pro(9)-Mammalian GnRH, Mammalian GnRH-ethylamide, Chicken II GnRH, Chicken II-ethylamide, D-Arg(6)-
15 Chicken II-ethylamide, D-Arg(6)-Chicken II-aza-Gly(10)-amide, Chicken I GnRH, Salmon GnRH, D-Arg(6)-Salmon GnRH-aza-Gly(10)-amide, and Lamprey GnRH was studied.

Mammalian GnRH was actively degraded by chorionic post-proline peptidase. While replacement of Gly(10)-NH₂ with ethylamide made each of the
20 mammalian GnRH analogs more resistant to degradation than Mammalian GnRH alone, some of these Mammalian GnRH were still degraded by the post-proline peptidase. Of four mammalian GnRH ethylamides studied, des-Gly(10)-GnRH ethylamide, des-Gly(10), D-Trp(6)-GnRH ethylamide, des-Gly(10)-D-Leu(6)-GnRH ethylamide, and Buserilin, each competitively inhibited the degradation of
25 mammalian GnRH; thus they were degraded by the post-proline peptidase. The effect of des-Gly(10) GnRH on the degradation of mammalian GnRH by chorionic post-proline peptidase is shown in Figure 2. The less an analog is capable of competing with the GnRH for the post-proline peptidase, the more resistant it is to degradation by post-proline peptidase and the more stable the analog will be in the
30 tumor tissue and/or in the blood. Thus the existing mammalian GnRH analogs

commonly used in medicine can be degraded in tumor tissues.

This activity of chorionic post-proline peptidase was inhibited by OH-Pro(9)-GnRH, Lamprey GnRH, Chicken I GnRH, Antide, Chicken II GnRH, and Salmon GnRH with a relative potency of 1.5, 1.5, 0.6, 0.6, 0.2, and 0.2, respectively, compared to that for GnRH. In viewing this data, the OH-Pro(9)-GnRH and Lamprey GnRH were determined to be the best competitors for GnRH degradation by chorionic post-proline peptidase. (See Figure 6 for inhibitor constants for analogs of GnRH.) They are as or even more potent than mammalian GnRH. Antide and Chicken I GnRH are three fold less potent than GnRH, but two fold more potent than the Salmon GnRH or Chicken II GnRH. The addition of the ethylamide to mammalian GnRH, both with and without the D-Trp(6)-, D-Phe(6) substitution, decreased the competition with mammalian GnRH for chorionic post-proline peptidase degradation, but not as markedly as did the Im-btl-D-His(6) or Chicken II GnRH analogs. Both D-Arg(6)-Chicken II GnRH-aza-Gly(10)-amide and Im-btl-D-His(6)-GnRH-ethylamide were essentially inactive, i.e. <0.005 inhibitory activity for GnRH. Essentially these latter two GnRH's were greater than 200 fold less active in the inhibition of GnRH degradation by chorionic post-proline peptidase. Thus these analogs appear to be very stable in the presence of post proline peptidase activity, however the Im-btl-His(6) analog has reduced receptor potency. The stability of the D-Arg(6)-Chicken II GnRH-aza-Gly(10)-amide was found to not only be greater than 200 fold more stable than GnRH but it still has increased receptor potency. The action of D-Arg(6)-Chicken II GnRH-aza-Gly(10)-amide on the degradation of mammalian GnRH by chorionic post-proline peptidase is shown in Figure 3. It is believed by Applicants that the same or similar results could be obtained using non-mammalian GnRH isomers or analogs with similar amino acid structure as Chicken II GnRH such as but not limited to Herring GnRH, Dogfish GnRH, or Catfish GnRH.

Since chorionic post-proline peptidase is a peptidase with high specificity for the degradation of GnRH at the proline-glycine peptide bond it can also degrade other GnRH species containing the same bond. The synthetic mammalian GnRH

analogs such as Antide are degraded with reduced activity while other analogs such as D-Arg(6)-Chicken II GnRH-aza-Gly(10)-amide are resistant to degradation by this endogenous chorionic enzyme. Such a resistant analog can be useful in the regulation of tumor tissue GnRH activity.

5 Degradation of mammalian GnRH by the tumor tissue cells was essentially 100% after overnight incubation. Specific inhibitors of post-proline peptidase were used to demonstrate this activity in the tumor cell extracts. The degradation of mammalian GnRH was inhibited by bacitracin, but not EDTA, demonstrating the enzyme similarity to chorionic post proline peptidase. From this study it was found
10 that the aza-Gly(10)amide derivatives of Chicken II GnRH and Salmon GnRH have little if any degradation as compared to mammalian GnRH. Each Chicken II and its analogs were more stable than the mammalian GnRH analogs analyzed. It is believed by Applicants that the same or similar results could be obtained using non-mammalian GnRH isomers or analogs with similar amino acid structure as Chicken
15 II GnRH such as but not limited to Herring GnRH, Dogfish GnRH, or Catfish GnRH.

Although the enzyme competition system had already been developed, newly synthesized non-mammalian GnRH analogs have not been utilized in this system. Previous data generated by Applicants have demonstrated that the antiserum is specific for mammalian GnRH, thus reducing potential for cross-reaction of non-mammalian GnRH isoforms or its analogs in the assay used in these studies.
20

Biological Activity Studies

The tumor growth inhibiting activity of the non-mammalian GnRH analogs was studied. Such data can be used to determine biological activity including regulation of tumor cell growth, tumor proliferation, and tumor regression. Bio-potency was
25 studied by determining cell death and tumor regression. Thus a primary parameter of interest was indicating the cell viability in the tumor cells being regulated by the exogenous GnRH activities which were studied.

The biological activity of the newly synthesized non-mammalian GnRH analogs was studied using an in vitro human tumor cell culture system. This system allows
30 for replicated extended activity studies. Mammalian GnRH action on the tumor

tissue cell has been studied using a similar system. Applicants studied replicate cultures, thus allowing for comparison of different doses of each non-mammalian GnRH analog to mammalian GnRH. In these studies, the action of the most stable and receptor-active non-mammalian GnRH analogs on tumor cell viability were determined.

The bio-potency studies were done with a MCF-7 breast cancer cell culture system and the cell viability as a measure of survival was assessed using the Alamar Blue assay. The percent difference in the Alamar Blue optical density (OD) readings at 570 and 600 nm in the treated and untreated controls was determined.

These studies were done using mammalian GnRH, chicken II GnRH, Leuprolide, Buserelin, the D-Arg(6)-Chicken II GnRH-aza-Gly(10)-amide analog as well as the D-Leu(6)-Chicken II GnRH-aza-Gly(10)-amide analog. A dose-response study in quadruplicate cultures was performed. Cell viability was assessed after 24 and 48 hours of incubation with the activity agent. The data analysis of these tumor cell culture sets at 24 hours is shown in Figure 5. More specifically, Figure 5 illustrates the anti-proliferative, tumor regression activity of D-Arg(6)-Chicken II GnRH-aza-Gly(10)-amide as compared to controls and other isoforms and analogs of GnRH after 24 hours of incubation. In this Figure 5, A1 is medium 199 (no vehicle); A2 is medium 199 (with vehicle); B1-B3 is Leuprolide; C1-C3 is mammalian GnRH; D1-D3 is Chicken II GnRH; E1-E3 is D-Arg(6)-Chicken II GnRH-aza-Gly(10) amide; G1-G3 is Buserelin.

After 24 hours of incubation, an inhibition of cell proliferation was observed with the Chicken II GnRH and its analogs, while even the highest doses of mammalian GnRH analogs, Leuprolide, and Buserelin were totally inactive. (See Figure 5). The lowest dose of Chicken II studied (10^{-8} M) was more effective than 10^{-5} M mammalian GnRH. The D-Arg(6)-Chicken II GnRH-aza-Gly(10)-amide resulted in positive dose-related activity, which was markedly active at 10^{-5} M. After 48 hours of incubation this analog was equally as potent as at 24 hours, while its natural isoform lost potency due to degradation. The mammalian GnRH and its analogs were totally ineffective in the inhibition of the MCF-7 breast cancer cell

proliferation after 48 hours of continued exposure. These data demonstrate that D-Arg(6)-Chicken II GnRH-aza-Gly(10)-amide analog is a very stable and bioactive molecule in the regulation of tumor cell growth in the human MCF-7 breast cancer cells. It is believed by Applicants that the same or similar results could be obtained
5 using non-mammalian GnRH isomers or analogs with similar amino acid structure to Chicken II GnRH such as but not limited to Salmon GnRH, Herring GnRH, Dogfish GnRH, or Catfish GnRH.

Using an in vitro system to define bio-potency is expected to be predictive of in vivo activity. In addition to tumor cell action, since these newly synthesized
10 non-mammalian GnRH analogs are known to act directly at the placenta to inhibit steroidogenesis, these analogs are expected to be active at the ovarian level to inhibit steroidogenesis. This would act as an added benefit in the cancer therapy.

Methods for Regulating Tumor Cell Growth and Proliferation In Vivo

In vivo trials utilizing the non-mammalian GnRH analogs of the present
15 invention may be performed to inhibit tumor cell growth and proliferation to thus induce regression of cancer cells in a mammal. The mammal can include a human with cancer. As a proposed dose regimen, it is anticipated that a human between 100 lbs. and 250 lbs. be administered about 10 nanograms to 1.0 gram of a chicken II GnRH analog, salmon GnRH analog, or other non-mammalian GnRH
20 analog with similar amino acid structure. This would be expected to be effective for inhibiting tumor growth or metastasis in the mammal once administered.

It is envisioned that these non-mammalian GnRH analogs will be administered intra-nasally, orally, intramuscularly, transdermally or vaginally. However, virtually any mode of administration may be used in the practice of the invention. Treatment
25 with these analogs may require short-term, repeated administrations of the active non-mammalian GnRH analog or long-term continuous therapy until tumor regression has occurred. Repeated administration could be used as needed.

Numerous in vitro fertilization (IVF) protocols now routinely use mammalian GnRH analogs for ovulation timing and have been shown to be nontoxic, even after
30 weeks of administration. Long-term therapies with mammalian GnRH analogs have

been used for treatment of endometriosis, prostate cancer and other cancers and have been shown to be nontoxic, even after months of administration. Long-term therapies with mammalian GnRH analogs have been associated with a hypoestrogenic state, which is frequently a desired condition in certain cancer therapies. The effect on the pituitary GnRH receptor is expected to be minimal with these non-mammalian GnRH analogs and with this short duration of treatment. Thus, the specific receptor activity of these analogs makes it less likely to interfere with normal physiology.

In some trials, the dosing regimen can comprise a pulsatile administration of the analog over a 24-hour period, wherein the daily dosage is administered in relatively equal 1/24th fractions. For example, where the daily dose is about 2.4 micrograms, the patient would be administered about 0.1 micrograms per hour over a 24-hour period. Such a daily pulsatile administration would create a hormonal environment in the patient sufficient to inhibit tumor cell growth and proliferation and/or induce its regression. The particular pharmaceutical preparations may be created by one of skill in the pharmaceutical arts. Remington's Pharmaceutical Sciences Remington: The Science and Practice of Pharmacy, 19th edition, Vol. 102, A. R. Gennaro, ed., Mack Publishing co. Easton, PA. (1995), is specifically incorporated herein by reference for this purpose.

ANTIBODIES SPECIFIC FOR NON-MAMMALIAN GNRH

Another embodiment of the present invention is to utilize non-mammalian GnRH analogs to prepare antibodies that preferentially bind the non-mammalian GnRH peptide sequences, or that bind the tumor tissues or any other non-pituitary GnRH peptide or protein. It is anticipated that these non-mammalian GnRH antibodies may be used in a variety of screening assays. For example, these antibodies may be used to determine levels of GnRH are present in a sample as an indicator molecule. The levels of such GnRH may be used to monitor and follow a patient's tumor activity or growth, as well as an indicator of the tumor's presence. The antibodies to non-mammalian GnRH may be monoclonal or polyclonal antibodies.

Polyclonal antibodies may be created by standard immunization techniques, wherein the immunogen used will be the non-mammalian chicken II GnRH, salmon GnRH, herring GnRH analog, or the naturally occurring decapeptide of any of these described herein, or any other non-mammalian GnRH analog with similar amino acid structure. These peptides may be used either alone or together in a pharmaceutically acceptable adjuvant. The animal, such as a rabbit, would be administered several doses of the decapeptide preparation, and the levels of the animal's antibody blood levels monitored until an acceptable antibody level (titer) had been reached.

For the preparation of monoclonal antibodies, one would follow standard techniques for the immunization of an animal, again using the decapeptide non-mammalian GnRH peptide or its analog. Once sufficiently high acceptable antibodies are reached (titer) in the animal, the spleen of the animal would be harvested and then fused with an immortalized cell line, such as a cancer cell line, to produce a population of hybridoma cells. This hybridoma population of cells would then be screened for those cells that produce the highest amount of antibody that specifically binds the non-mammalian GnRH analog decapeptide. Such hybridoma cells would be selected, and then cultured. The antibody to non-mammalian GnRH would then be collected from the media of the cell culture using techniques well known to those of skill in the art.

For purposes of the practice of preparing polyclonal and monoclonal antibody, the textbook Sambrook et al (1989) *Molecular Cloning, A Laboratory Manual*, 2nd Ed., Cold Springs Harbor Laboratory, Cold Springs Harbor, N.Y., is specifically incorporated herein by reference. In addition to the embodiments presented it is believed by Applicants that the disclosed non-mammalian GnRH isomers and/or analogs as well as any other gene regulator can be used to regulate the gene expression of non-mammalian GnRH or expression of its receptors in tumor cells. It is further believed by Applicants that the non-mammalian GnRH analogs disclosed can be used in the development of stable, toxin conjugated antibodies or ligands that can specifically bind to the GnRH receptor on the tumor cell and kill the

cell.

All of the compositions and methods disclosed and claimed herein can be made and executed without undue experimentation in light of the present disclosure. While the compositions and methods of this invention have been
5 described in terms of preferred embodiments, it will be apparent to those of skill in the art that variations may be applied to the composition, methods and in the steps or in the sequence of steps of the method described therein without departing from the concept, spirit and scope of the invention. More specifically, it will be apparent that certain agents, who are both chemically and physiologically, related, might be
10 substituted for the agents described herein while the same or similar results would be achieved. All such similar substitutes and modifications apparent to those skilled in the art are deemed to be within the spirit, scope and concept of the invention as defined by the appended claims.

REFERENCES

1. Burgus R, Guillemin R 1970 Hypothalamic releasing factors. *Ann Rev Biochem* 39:499-526
2. Baba Y, Matsuo H, Schally AV 1971 Structure of the porcine LH- and
5 FSH-releasing hormone. II. Confirmation of the proposed structure by conventional sequential analyses. *Biochem Biophys Res Commun* 44:459-463
3. Corbin A 1982 From contraception to cancer: A review of the therapeutic applications of LHRH analogues as antitumor agents. *Yale J Biol Med* 55:27-47
4. Buzdar AU, Hortobagyi G 1998 Update on endocrine therapy for breast cancer.
10 *Clin Cancer Res* 4:527-534
5. Bare, R.L. and F.M. Torti. 1998. Endocrine therapy of prostate cancer. In *Biological and hormonal therapies of cancer*. K.A. Foon and H.B. Muss, editors. Kluwer Academic Publishers, Boston. 69-86.
6. Siler-Khodr TM, Khodr GS 1978 Luteinizing hormone releasing factor content
15 of the human placenta. *Am J Obstet Gynecol* 130:216-219
7. Khodr GS, Siler-Khodr TM 1978 Localization of luteinizing hormone releasing factor (LRF) in the human placenta. *Fert Steril* 29:523-526
8. Siler-Khodr TM, Khodr GS 1979 Extrahypothalamic luteinizing hormone releasing factor (LRF): Release of immunoreactive LRF by the human placenta in
20 vitro. *Fert Steril* 22:294-296
9. Khodr GS, Siler-Khodr TM 1980 Placental LRF and its synthesis. *Science* 207:315-317
10. Siler-Khodr, T.M. 1992. The Placenta: Part IV-Function of the Human Placenta. In *Neonatal and Fetal Medicine*. R.A. Polin and W.W. Fox, editors. W.B.
25 Saunders Co. Philadelphia, PA. 74-86.
11. Youngblood WW, Humm J, Kizer JS 1979 TRH-like immunoreactivity in rat pancreas and eye, bovine and sheep pineals, and human placenta: Non-identity with synthetic Pyroglu-His-Pro-NH₂ (TRH). *Brain Res* 163:101-110
12. Dubois MP 1975 Immunoreactive somatostatin is present in discrete cells of
30 the endocrine pancreas. *Proc Natl Acad Sci USA* 72:1340-1343

13. Adashi, E.Y. 1996. The Ovarian Follicular Apparatus. In Lippincott-Raven Publishers. E.Y. Adashi, J.A. Rock, and Z. Rosenwaks, editors. Lippincott-Raven Publishers, Philadelphia. 17-40.
14. Szende B, Srkalovic G, Groot K, Lapis K, Schally AV 1990 Growth inhibition
5 of mouse MXT mammary tumor by the luteinizing hormone-releasing hormone antagonist SB-75. J Natl Cancer Inst 82:513-517
15. Srkalovic G, Wittliff JL, Schally AV 1990 Detection and partial characterization of receptors of [D-Trp(6)]-luteinizing hormone-releasing hormone and epidermal growth factor in human endometrial carcinoma. Cancer Res 50:1841-1846
- 10 16. Szende B, Srkalovic G, Groot K, Lapis K, Schally AV 1991 Regression of nitrosamine-induced pancreatic cancers in hamsters treated with luteinizing hormone-releasing hormone antagonists or agonists. Cancer Res 50:3716-3721
- 15 17. Ohno T, Atsushi I, Furui T, Takahashi K, Tamaya T 1993 Presence of gonadotropin-releasing hormone and its messenger ribonucleic acid in human ovarian epithelial carcinoma. Am J Obstet Gynecol 169:605-610
18. Palyi I, Vincze B, Kalnay A, Turi G, Mezo I, Teplan I, Seprodi J, Pato J, Mora M 1996 Effect of gonadotropin-releasing hormone analogs and their conjugates on gonadotropin-releasing hormone receptor-positive human cancer cell lines. Cancer Detect Prev 20:146-152
- 20 19. Teissmann T, Klenner T, Deger W, Hilgard P, McGregor GP, Voigt K, Engel J 1996 Pharmacological studies with cetrorelix (SB-75), a potent antagonist of luteinising hormone-releasing hormone. Eur J Cancer 32A:1574-1579
20. Chatzaki E, Bax MR, Eidne KA, Anderson L, Grudzinskas JG, Gallagher CJ 1996 The expression of gonadodtrophin-releasing hormone and its receptor in
25 endometrial cancer, and its relevance as an autocrine growth factor. Cancer Res 56:2059-2065
21. Jungwirht A, Galvan G, Pinski J, Halmos G, Szepeshazi K, Cai RZ, Groot K, Schally AV 1997 Luteinizing hormone-releasing hormone antagonist cetrorelix (SB-75) and bombesin antagonist RC-3940-II inhibit the growth of
30 androgen-independent PC-3 prostate cancer in nude mice. Prostate 32:164-172

22. Jungwirth A, Pinski J, Galvan G, Halmos G, Szepeshazi K, Cai RZ, Groot K, Vadillo-Buenfil M, Schally AV 1997 Inhibition of growth of androgen-independent DU-145 prostate cancer in vivo by luteinising hormone-releasing hormone antagonist cetorelix and bombesin antagonists RC-3940-II and RC-3950-II. Eur J
5 Cancer 33:1141-1148
23. Bahk JY, Hyun JS, Lee H, Kim MO, Cho GJ, Lee BH, Choi WS 1998 Expression of gonadotropin-releasing hormone (GnRH) and GnRH receptor mRNA in prostate cancer cells and effect of GnRH on the proliferation of prostate cancer cells. Urol Res 26:259-264
- 10 24. Van Groeninghen JC, Kiesel L, Winkler D, Zwirner M 1998 Effect of luteinising-hormone-releasing hormone on nervous-system tumors. Lancet 352:372-373
25. Yin H, Cheng KW, Hwa H, Peng C, Auersperg N, Leung PCK 1998 Expression of the messenger RNA for gonadotropin-releasing hormone and its receptor in
15 human cancer cell lines. Life Sci 62:2015-2023
26. Lamharzi N, Schally AV, Koppan M 1998 Luteinizing hormone-releasing hormone (LH-RH) antagonist Cetorelix inhibits growth of DU-145 human androgen-independent prostate carcinoma in nude mice and suppresses the levels and mRNA expression of IGF-II in tumors. Regul Pept 77:185-192
- 20 27. Brewer CA, Shevlin D 1998 Encouraging response of an advanced steroid-cell tumor to GnRH agonist therapy. Obstet Gynecol 92:661-663
28. Mesia AF, Williams FS, Yan Z, Mittal K 1998 Aborted leiomyosarcoma after treatment with leuprolide acetate. Obstet Gynecol 92:664-666
29. Motomura S 1998 Inductions of apoptosis in ovarian carcinoma cell line by
25 gonadotropin-releasing hormone agonist. Kurume Med J 45:27-32
30. Imai A, Takagi A, Horibe S, Takagi H, Tamaya T 1998 Fas and Fas ligand system may mediate antiproliferative activity of gonadotropin-releasing hormone receptor in endometrial cancer cells. Int J Oncol 13:97-100
31. Lamharzi N, Halmos G, Jungwirth A, Schally AV 1998 Decrease in the level
30 and mRNA expression of LH-RH and EGF receptors after treatment with LH-RH

anatagonist Cetrorelix in DU-145 prostate tumor xenografts in nude mice. *Int J Oncol* 13:429-435

32. Sherwood NM, Lovejoy DA, Coe IR 1993 Origin of mammalian gonadotropin-releasing hormones. *Endocr Rev* 14:241-254

5 33. King JA, Millar RP 1995 Evolutionary aspects of gonadotropin-releasing hormone and its receptor. *Cell Mol Neurobiol* 15:5-23

34. Siler-Khodr TM, Khodr GS, Valenzuela G 1984 Immunoreactive gonadotropin-releasing hormone level in maternal circulation throughout pregnancy. *Am J Obstet Gynecol* 150:376-379

10 35. Sorem KA, Smikle CB, Spencer DK, Yoder BA, Grayson MA, Siler-Khodr TM 1996 Circulating maternal CRH and GnRH in normal and abnormal pregnancies. *Am J Obstet Gynecol* 175:912-916

36. Khodr GS, Siler-Khodr TM 1979 The effect of luteinizing hormone releasing factor (LRF) on hCG secretion. *Fert Steril* 30:301-304

15 37. Siler-Khodr TM, Khodr GS 1981 Dose response analysis of GnRH stimulation of hCG release from human term placenta. *Biol Reprod* 25:353-358

38. Siler-Khodr TM, Khodr GS, Valenzuela G, Rhode J 1986 Gonadotropin-releasing hormone effects on placental hormones during gestation: I. Alpha-human chorionic gonadotropin, human chorionic gonadotropin and human
20 chorionic somatomammotropin. *Biol Reprod* 34:245-254

39. Siler-Khodr TM, Khodr GS, Valenzuela G, Harper MJ, Rhode J 1986 GnRH effects on placental hormones during gestation. III. Prostaglandin E, prostaglandin F, and 13,14-dihydro-15-keto-prostaglandin F. *Biol Reprod* 35:312-319

40. Kang IS, Koong MK, Forman JS, Siler-Khodr TM 1991 Dose-related action of
25 GnRH on basal prostanoid production from the human term placenta. The 38th Annual Meeting of the Society for Gynecologic Investigation (San Antonio) Abstract #310:253(Abstr.)

41. Siler-Khodr TM, Khodr GS, Harper MJ, Rhode J, Vickery BH, Nestor JJ, Jr. 1986 Differential inhibition of human placental prostaglandin release in vitro by a
30 GnRH antagonist. *Prostaglandins* 31:1003-1010

42. Siler-Khodr, T.M. and G.S. Khodr. 1981. The production and activity of placental releasing hormones. In *Fetal Endocrinology*. J. Resko and W. Montagna, editors. Academic Press, Inc. New York. 183-210.
43. Siler-Khodr, T.M. and G.S. Khodr. 1982. GnRH in the placenta. In *Role of Peptides and Proteins in Control of Reproduction*. D.S. Dhindsa and S.M. McCann, editors. Elsevier North Holland, New York. 347-363.
44. Siler-Khodr TM 1983 Hypothalamic-like releasing hormones of the placenta. *Clin Perinatol* 10:553-566
45. Siler-Khodr TM 1983 Hypothalamic-like peptides of the placenta. *Semin Reprod Endocrinol* 1:321-333
46. Shi LY, Zhang ZW, Li WX 1994 Regulation of human chorionic gonadotropin secretion and messenger ribonucleic acid levels by follistatin in the NUCC-3 choriocarcinoma cell line. *Endocrinology* 134:2431-2437
47. Steele GL, Currie WD, Yuen BH, Jia XC, Perlas E, Leung PC 1993 Acute stimulation of human chorionic gonadotropin secretion by recombinant human activin-A in first trimester human trophoblast. *Endocrinology* 133:297-303
48. Li W, Olofsson JI, Jeung EB, Krisinger J, Yuen BH, Leung PC 1994 Gonadotropin-releasing hormone (GnRH) and cyclic AMP positively regulate inhibin subunit messenger RNA levels in human placental cells. *Life Sci* 55:1717-1724
49. Petraglia F, Vaughan J, Vale W 1991 Inhibin and activin modulate the release of gonadotropin-releasing hormone, human chorionic gonadotropin, and progesterone from cultured human placental cells. *Proc Natl Acad Sci USA* 86:5114-5117
50. Petraglia F, Sawchenko P, Lim ATW, Rivier J, Vale W 1987 Localization, secretion, and action of inhibin in human placenta. *Science* 237:187-189
51. Shi CZ, Zhuang LZ 1993 Norepinephrine regulates human chorionic gonadotrophin production by first trimester trophoblast tissue in vitro. *Placenta* 14:683-693
52. Cemerikic B, Maulik D, Ahmed MS 1992 Opioids regulation of hCG release from trophoblast tissue is mediated by LHRH. *Placenta Abstract*:9(Abstr.)

53. Petraglia F, Vaughan J, Vale W 1990 Steroid hormones modulate the release of immunoreactive gonadotropin-releasing hormone from cultured human placental cells. *J Clin Endocrinol Metab* 70:1173-1178
54. Haning RV, Jr., Choi L, Kiggins AJ, Kuzma DL, Summerville JW 1982 Effects of dibutyladenosine 3',5'-monophosphate, luteinizing hormone-releasing hormone, and aromatase inhibitor on simultaneous outputs of progesterone 17 β -estradiol, and human chorionic gonadotropin by term placental explants. *J Clin Endocrinol Metab* 55:213-218
55. Petraglia F, Lim AT, Vale W 1987 Adenosine 3',5'-monophosphate, prostaglandins, and epinephrine stimulate the secretion of immunoreactive gonadotropin-releasing hormone from cultured human placental cells. *J Clin Endocrinol Metab* 65:1020-1025
56. Haning RV, Jr., Choi L, Kiggins AJ, Kuzma DL 1982 Effects of prostaglandins, dibutyl camp LHRH, estrogens, progesterone, and potassium on output of prostaglandin F2a, 13,14-dihydro-15-keto-prostaglandin F2a, hCG, estradiol, and progesterone by placental minces. *Prostaglandins* 24:495-506
57. Barnea ER, Feldman D, Kaplan M 1991 The effect of progesterone upon first trimester trophoblastic cell differentiation and human chorionic gonadotrophin secretion. *Hum Reprod* 6:905-909
58. Barnea ER, Kaplan M 1989 Spontaneous, gonadotropin-releasing hormone-induced, and progesterone-inhibited pulsatile secretion of human chorionic gonadotropin in the first trimester placenta in vitro. *J Clin Endocrinol Metab* 69:215-217
59. Branchaud C, Goodyer C, Lipowski L 1983 Progesterone and estrogen production by placental monolayer cultures: Effect of dehydroepiandrosterone and luteinizing hormone-releasing hormone. *J Clin Endocrinol Metab* 56:761-766
60. Ahmed NA, Murphy BE 1988 The effects of various hormones on human chorionic gonadotropin production in early and late placental explant cultures. *Am J Obstet Gynecol* 159:1220-1227

61. Iwashita M, Watanabe M, Adachi T, Ohira A, Shinozaki Y, Takeda Y, Sakamoto S 1989 Effect of gonadal steroids on gonadotropin-releasing hormones stimulated human chorionic gonadotropin release by trophoblast cells. *Placenta* 10:103-112
62. Haning RV, Jr., Choi L, Kiggins AJ, Kuzma DL, Summerville JW 1982 Effects of dibutyl cAMP, LHRH, and aromatase inhibitor on simultaneous outputs of prostaglandin F2a, and 13,14-dihydro-15-keto-prostaglandin F2a by term placental explants. *Prostaglandins* 23:29-40
63. Wilson E, Jawad M 1980 Luteinizing hormone-releasing hormone suppression of human placental progesterone production. *Fert Steril* 33:91-93
- 10 64. Duello TM, Tsai SJ, Van Ess PJ 1993 In situ demonstration and characterization of progonadotropin-releasing hormone messenger ribonucleic acid in first trimester human placentas. *Endocrinology* 133:2617-2623
65. Kelly AC, Rodgers A, Dong KW, Barrezueta NX, Blum M, Roberts JL 1991 Gonadotropin-releasing hormone and chorionic gonadotropin gene expression in human placental development. *DNA Cell Biol* 10:411-421
- 15 66. Berry SA, Pescovitz OH 1988 Identification of a rat GHRH-like substance and its messenger RNA in rat testis. *Endocrinology* 123:661-663
67. Radovick S, Wondisford FE, Nakayama Y, Yamada M, Cutler GB, Jr., Weintraub BD 1990 Isolation and characterization of the human gonadotropin-releasing hormone gene in the hypothalamus and placenta. *Mol Endocrinol* 4:476-480
- 20 68. Adelman JP, Mason AJ, Hayflick JS, Seeburg PH 1986 Isolation of the gene and hypothalamic cDNA for the common precursor of gonadotropin-releasing hormone and prolactin release-inhibiting factor in human and rat. *Proc Natl Acad Sci USA* 83:179-183
- 25 69. Rakoff J, VandenBerg G, Siler TM, Yen SSC 1973 An integrated direct functional test of the adenohipophysis. The Pacific Coast Obstetrics and Fertility Society, Las Vegas (Abstract):(Abstr.)
70. Dong KW, Yu KL, Roberts JL 1993 Identification of a major up-stream transcription start site for the human progonadotropin-releasing hormone gene used
- 30

in reproductive tissues and cell lines. *Mol Endocrinol* 7:1654-1666

71. Dong KW, Duval P, Zeng Z, Gordon K, Williams RF, Hodgen GD, Jones G, Kerdelhue B, Roberts JL 1996 Multiple transcription start sites for the GnRH gene in rhesus and cynomolgus monkeys: a non-human primate model for studying
5 GnRH gene regulation. *Mol Cell Endocrinol* 117:121-130

72. Dong KW, Yu KL, Chen ZG, Chen YD, Roberts JL 1997 Characterization of multiple promoters directing tissue-specific expression of the human gonadotropin-releasing hormone gene. *Endocrinology* 138:2754-2762

73. Chandran UR, Attardi B, Friedman R, Dong KW, Roberts JL, DeFranco DB
10 1994 Glucocorticoid receptor-mediated repression of gonadotropin-releasing hormone promoter activity in GT1 hypothalamic cell lines. *Endocrinology* 134:1467-1474

74. Dong KW, Chen ZG, Cheng KW, Yu KL 1996 Evidence for estrogen receptor-mediated regulation of human gonadotropin-releasing hormone promoter
15 activity in human placental cells. *Mol Cell Endocrinol* 117:241-246

75. Joss JM, King JA, Millar RP 1994 Identification of the molecular forms of and steroid hormone response to gonadotropin-releasing hormone in the Australian lungfish, *Neoceratodus forsteri*. *Gen Comp Endocrinol* 96:392-400

76. Montero M, Le Belle N, King JA, Millar RP, Dufour S 1995 Differential
20 regulation of the two forms of gonadotropin-releasing hormone (mGnRH and cGnRH-II) by sex steroids in the European female silver eel (*Anguilla anguilla*). *Neuroendocrinology* 61:525-535

77. Ikeda M, Taga M, Sakakibara H, Minaguchi H, Ginsburg E, Vonderhaar BK
25 1996 Gene expression of gonadotropin-releasing hormone in early pregnant rat and steroid hormone exposed mouse uteri. *J Endocrinol Invest* 19:708-713

78. Gothilf Y, Meiri I, Elizur A, Zohar Y 1997 Preovulatory changes in the levels of three gonadotropin-releasing hormone-encoding messenger ribonucleic acids (mRNAs), gonadotropin B-subunit mRNAs plasma gonadotropin, and steroids in the female gilthead seabream, *Sparus aurata*. *Biol Reprod* 57:1145-1154

30 79. Bramley TA, McPhie CA, Menzies GS 1994 Human placental

gonadotrophin-releasing hormone (GnRH) binding sites: III. Changes in GnRH binding levels with stage of gestation. *Placenta* 15:733-745

80. Lin LS, Roberts VJ, Yen SS 1997 Expression of human gonadotropin-releasing hormone receptor gene in the placenta and its functional relationship to human chorionic gonadotropin secretion. *J Clin Endocrinol Metab* 80:580-585

81. Szilagyi A, Benz R, Rossmanith WG 1992 The human first-term placenta in vitro: regulation of hCG secretion by GnRH and its antagonist. *Gynecol Endocrinol* 6:293-300

82. Currie WD, Setoyama T, Lee PS, Baimbridge KG, Church J, Yuen BH, Leung PC 1993 Cytosolic free Ca^{2+} in human syncytiotrophoblast cells increased by gonadotropin-releasing hormone. *Endocrinology* 133:2220-2226

83. Emons G, Muller V, Ortmann O, Schulz KD 1998 Effects of LHRH-analogues on mitogenic signal transduction in cancer cells. *J Steroid Biochem Mol Biol* 65:1-6

84. Pahwa GS, Kullander S, Vollmer G, Oberheuser F, Knuppen R, Emons G 1991 Specific low affinity binding sites for gonadotropin releasing hormone in human endometrial carcinoma. *Eur J Obstet Gynecol Reprod Biol* 41:135-142

85. Nagy A, Schally AV, Armatis P, Szepeshazi K, Halmos G, Kovacs M, Zarandi M, Groot K, Miyazaki M, Jungwirth A, Horvath JE 1996 Cytotoxic analogs of luteinizing hormone-releasing hormone containing doxorubicin or 2-pyrrolinodoxorubicin, a derivative 500-1000 times more potent. *Proc Natl Acad Sci USA* 93:7269-7273

86. Vincze B, Palyi I, Gaal D, Pato J, Mora M, Mezo I, Teplan I, Sepródi J 1996 In vivo studies of the new gonadotropin-releasing hormone antagonist-copolymer conjugates having antitumor activity. *Cancer Detect Prev* 20:153-159

87. Neri C, Berthois Y, Schatz B, Drieu K, Martin PM 1990 Compared effects of GnRH analogs and 4-hydroxytamoxifen on growth and steroid receptors in antiestrogen sensitive and resistant MCF-7 breast cancer cell sublines. *Breast Cancer Res Treat* 15:85-93

88. Crighton IL, Dowsett M, Lal A, Man A, Smith IE 1989 Use of luteinising hormone-releasing hormone agonist (Leuporelin) in advanced post-menopausal

breast cancer. *Br J Cancer* 60:644-648

89. Teodorczyk-Injeyan J, Jewett MAS, Kellen JA, Malkin A 1981 Gonadoliberin (LHRH) mediated release of choriogonadotropin in experimental human and animal tumors in vitro. *Endocr Res Commun* 8:19-24

5 90. Bruckner HW, Motwani BT 1989 Treatment of advanced refractory ovarian carcinoma with a gonadotropin-releasing hormone analogue. *Clin Med* 161:1216-1218

91. Cassano A, Astone A, Garufi C, Noviello MR, Pietrantonio F, Barone C 1987 A response in advanced post-menopausal breast cancer during treatment with the luteinising hormone releasing hormone agonist-Zoladex. *Exp Biol Med* 48:123-124

10 92. Klijn JGM, DeJong FH 1982 Treatment with a luteinising-hormone releasing-hormone analogue (Busereline) in premenopausal patients with metastatic breast cancer. *Lancet* 1982:1213-1216

93. Sealton SC, Weinstein H, Millar RP 1997 Molecular mechanism of ligand interaction with the gonadotropin-releasing hormone receptor. *Endocr Rev* 15 18:180-205

94. Karten MJ, Rivier JE 1986 Gonadotropin-releasing hormone analog design. Structure-function studies toward the development of agonists and antagonists: Rationale and perspective. *Endocr Rev* 7:44-66

20 95. Kang IS, Koong MK, Forman JS, Siler-Khodr TM 1991 Dose-related action of gonadotropin-releasing hormone on basal prostanoid production from the human term placenta. *Am J Obstet Gynecol* 165:1771-1776

96. Gautron JP, Pattou E, Kordon C 1981 Occurrence of higher molecular forms of LHRH in fractionated extracts from rat hypothalamus, cortex and placenta. *Mol Cell Endocrinol* 24:1-15

25 97. Mathialagan N, Rao AJ 1986 Gonadotropin releasing hormone in first trimester human placenta: Isolation, partial characterisation and in vitro biosynthesis. *J Biosci* 10:429-441

98. Gautron JP, Pattou E, Bauer K, Rotten D, Kordon C 1989 LHRH-like immunoreactivity in the human placenta is not identical to LHRH. *Placenta* 30

10:19-35

99. Nowak RA, Wiseman BS, Bahr JM 1984 Identification of a gonadotropin-releasing hormone-like factor in the rabbit fetal placenta. *Biol Reprod* 31:67-75(Abstr.)
- 5 100. Siler-Khodr, T.M. 1987. LHRH in pregnancy. In *LHRH and Its Analogs: Contraceptive and Therapeutic Applications, Part II*. B.H. Vickery and J.J. Nestor, Jr. editors. MTP Press, Ltd. Lancaster. 161-178.
101. Siler-Khodr, T.M. 1988. Hypothalamic-like activities of fetal membranes. In *Proceedings of the 1st International Symposium on the Physiology of Human Fetal Membranes*. J. Challis and B. Mitchell, editors. Perinatology Press, Ithaca, NY. 91-116.
- 10 102. Siler-Khodr TM, Kang IS, Jones MA, Harper MJK, Khodr GS, Rhode J 1989 Characterization and purification of a placental protein that inactivates GnRH, TRH and Angiotensin II. *Placenta* 10:283-296
- 15 103. Kang IS, Siler-Khodr TM 1992 Chorionic peptidase inactivates GnRH as a post-proline peptidase. *Placenta* 13:81-87
104. Kelsall R, Coe IR, Sherwood NM 1990 Phylogeny and ontogeny of gonadotropin-releasing hormone: Comparison of guinea pig, rat, and a protochordate. *Gen Comp Endocrinol* 479-494
- 20 105. Powell JF, Reska-Skinner SM, Prakash MO, Fischer WH, Park M, Rivier JE, Craig AG, Mackie GO, Sherwood NM 1996 Two new forms of gonadotropin-releasing hormone in a protochordate and the evolutionary implications. *Proc Natl Acad Sci USA* 93:10461-10464
106. Carolsfeld J, Powell JFF, Park M, Fischer WH, Craig AG, Chang JP, Rivier JE, 25 Sherwood NM 2000 Primary structure and function of three gonadotropin-releasing hormones, including a novel form, from an ancient teleost, herring. *Endocrinology* 141:505-512
107. Powell JF, Zohar Y, Elizur A, Park M, Fischer WH, Craig AG, Rivier JE, Lovejoy DA, Sherwood NM 1994 Three forms of gonadotropin-releasing hormone 30 characterized from brains of one species. *Proc Natl Acad Sci USA* 91:12081-12085

108. Montero M, Vidal B, King JA, Tramu G, Vandesande F, Dufour S, Kah O 1994 Immunocytochemical localization of mammalian GnRH (gonadotropin-releasing hormone) and chicken GnRH-II in the brain of the European silver eel (*Anguilla anguilla* L.). *J Chem Neuroanat* 7:227-241
- 5 109. White SA, Kasten TL, Bond CT, Adelman JP, Fernald RD 1995 Three gonadotropin-releasing hormone genes in one organism suggest novel roles for an ancient peptide. *Proc Natl Acad Sci USA* 92:8363-8367
110. Powell JF, Fischer WH, Park M, Craig AG, Rivier JE, White SA, Francis RC, Fernald RD, Licht P, Warby C, et al 1995 Primary structure of solitary form of gonadotropin-releasing hormone (GnRH) in cichlid pituitary; three forms of GnRH
10 in brain of cichlid and pumpkinseed fish. *Regul Pept* 57:43-53
111. Zohar Y, Elizur A, Sherwood NM, Powell JF, Rivier JE, Zmora N 1995 Gonadotropin-releasing activities of the three native forms of gonadotropin-releasing hormone present in the brain of gilthead seabream, *Sparus aurata*. *Gen Comp*
15 *Endocrinol* 97:289-299
112. Lin XW, Peter RE 1996 Expression of salmon gonadotropin-releasing hormone (GnRH) and chicken GnRH-II precursor messenger ribonucleic acids in the brain and ovary of goldfish. *Gen Comp Endocrinol* 101:282-296
113. Di Fiore MM, King JA, D'Aniello B, Rastogi RK 1996 Immunoreactive
20 mammalian and chicken-II GnRHs in *Rana esculenta* brain during development. *Regul Pept* 62:119-124
114. Powell JF, Krueckl SL, Collins PM, Sherwood NM 1996 Molecular forms of GnRH in three model fishes: rockfish, medaka and zebrafish. *J Endocrinol* 150:17-23
- 25 115. Iela L, Powell JFF, Sherwood NM, D'Aniello B, Rastogi RK, Bagnara JT 1996 Reproduction in the Mexican leaf frog, *Pachymedusa dacnicolor*. VI. Presence and distribution of multiple GnRH forms in the brain. *Gen Comp Endocrinol* 103:235-243
116. Powell JF, Standen EM, Carolsfeld J, Borella MI, Gazola R, Fischer WH, Park
30 M, Craig AG, Warby CM, Rivier JE, Val-Sella MV, Sherwood NM 1997 Primary

- structure of three forms of gonadotropin-releasing hormone (GnRH) from the pacu brain. *Regul Pept* 68:189-195
117. Dellovade TL, King JA, Millar RP, Rissman EF 1993 Presence and differential distribution of distinct forms of immunoreactive gonadotropin-releasing hormone in
5 the musk shrew brain. *Neuroendocrinology* 58:166-177
118. King JA, Steneveld AA, Curlewis JD, Rissman EF, Millar RP 1994 Identification of chicken GnRH II in brains of metatherian and early-evolved eutherian species of mammals. *Regul Pept* 54:467-477
119. Kasten TL, White SA, Norton TT, Bond CT, Adelman JP, Fernald RD 1996
10 Characterization of two new preproGnRH mRNAs in the tree shrew: first direct evidence for mesencephalic GnRH gene expression in a placental mammal. *Gen Comp Endocrinol* 104:7-19
120. Jimenez-Linan M, Rubin BS, King JC 1997 Examination of guinea pig luteinizing hormone-releasing hormone gene reveals a unique decapeptide and
15 existence of two transcripts in the brain. *Endocrinology* 138:4123-4130
121. White SA, Bond CT, Francis RC, Kasten TL, Fernald RD, Adelman JP 1994 A second gene for gonadotropin-releasing hormone: cDNA and expression pattern in the brain. *Proc Natl Acad Sci USA* 91:1423-1427
122. Lin XW, Peter RE 1997 Cloning and expression pattern of a second
20 [His5Trp7Tyr8]gonadotropin-releasing hormone (chicken GnRH-H-II) mRNA in goldfish: evidence for two distinct genes. *Gen Comp Endocrinol* 107:262-272
123. Siler-Khodr TM, Grayson M 1999 Comparison of GnRH and its synthetic and naturally occurring analogs for binding to the human placental receptor. *J Soc Gynecol Invest* 6:225A(Abstr.)
124. Siler-Khodr TM, Grayson M 2000 Inhibition of human trophoblast function by
25 superagonists of chicken II GnRH. *J Soc Gynecol Invest* 7:280A(Abstr.)
125. Mezo I, Seprodi J, Vincze B, Palyi I, Keri G, Vadasz Z, Toth G, Kovacs M, Koppan M, Horvath JE, Kalnay A, Teplan I 1996 Synthesis of GnRH analogs having direct antitumor and low LH-releasing activity. *Biomedical Peptides, Proteins &*
30 *Nucleic Acids* 2:33-40

126. Mezo I, Lovas S, Palyi I, Vincze B, Kalnay A, Turi G, Vadasz Z, Seprodi J, Idei M, Toth G, Gulyas E, Otvos F, Mak M, Horvath JE, Teplan I, Murphy RF 1997 Synthesis of gonadotropin-releasing hormone III analogs. Structure-antitumor activity relationships. J Med Chem 40:3353-3358

WE CLAIM:

1. A composition comprising a non-mammalian GnRH analog for use in regulating tumor GnRH activity, wherein said analog is capable of binding to tumor GnRH receptors with greater affinity than mammalian GnRH and is active in the presence of a post-proline peptidase or endopeptidase.
2. The composition of Claim 1 wherein said non-mammalian GnRH analog has a D-amino acid substitution at position 6 and a post-proline peptidase inhibitor at position 10.
3. The composition of Claim 2 wherein said post-proline peptidase inhibitor is selected from the group consisting of aza-Gly-amide or ethylamide.
4. The composition of Claim 2 wherein said non-mammalian GnRH analog is further defined as an anti-tumor agent.
5. The composition of Claim 2 wherein said non-mammalian GnRH analog is further defined as an anti-proliferative agent.
6. The composition of Claim 5 wherein said non-mammalian GnRH analog is further defined as an anti-metastatic agent.
7. The composition of Claim 6 wherein said non-mammalian GnRH analog is further defined as an apoptotic agent.
8. The composition of Claim 2 wherein said non-mammalian GnRH analog is selected from the group consisting of Chicken II GnRH analog, Salmon GnRH analog, and Herring GnRH analog.
9. The composition of Claim 8 wherein said Chicken II GnRH analog is selected from the group consisting of D-Arg(6)-Chicken II GnRH-ethylamide and D-Arg(6)-Chicken II GnRH-aza-Gly(10)-amide.
10. The composition of Claim 8 wherein said Salmon GnRH analog is selected from the group consisting of D-Arg(6)-Salmon-GnRH-ethylamide and D-Arg(6)-Salmon-GnRH-aza-Gly(10)-amide.
11. The composition of Claim 9 wherein said D-Arg(6)-Chicken II GnRH-aza-Gly(10)-amide has a sequence as defined in SEQ ID NO. 2.
12. The composition of Claim 8 wherein said Chicken II GnRH has a cDNA

sequence of SEQ ID NO. 1.

13. The composition of Claim 10 wherein said D-Arg(6)-Salmon-GnRH-aza-Gly(10)-amide has a sequence as defined in SEQ ID NO. 4.

14. The composition of Claim 8 wherein said Salmon GnRH has a cDNA sequence
5 of SEQ ID NO. 3.

15. The composition of Claim 8 wherein said Herring GnRH analog has a sequence as defined in SEQ ID NO. 6.

16. The composition of Claim 8 wherein said Herring GnRH has a cDNA sequence of SEQ ID NO. 5.

10 17. The composition of Claim 2 wherein said D-amino acid substituted at position 6 is selected from a group consisting of D-Arg, D-Leu, D-tBu-Ser, and D-Trp.

18. A method for regulating tumor activity using a non-mammalian GnRH analog comprising the step of administering to a mammal a pharmaceutical preparation of the analog, wherein said analog is capable of binding to tumor GnRH receptors with
15 greater affinity than mammalian GnRH and is active in the presence of a post-proline peptidase or endopeptidase due to a D-amino acid substitution at position 6 and a post-proline peptidase inhibitor at position 10.

19. The method of Claim 18 wherein said non-mammalian GnRH analog is selected from the group consisting of Chicken II GnRH analog, Salmon GnRH
20 analog, and Herring GnRH analog.

20. The method of Claim 19 wherein said Chicken II GnRH analog is selected from the group consisting of D-Arg(6)-Chicken II GnRH-ethylamide and D-Arg(6)-Chicken II GnRH-aza-Gly(10)-amide.

21. The method of Claim 19 wherein said Salmon GnRH analog is selected from
25 the group consisting of D-Arg(6)-Salmon-GnRH-ethylamide and D-Arg(6)-Salmon-GnRH-aza-Gly(10)-amide.

SEQ ID NO: 1

Chicken II

cDNA

5 CAG CAC TGG TCT CAT GGC TGG TAT CCT GGA

SEQ ID NO: 2

Chicken II GnRH Analog

p-Glu-His-Trp-Ser-His-D-Arg-Trp-Tyr-Pro-aza-Gly-NH₂

10

SEQ ID NO: 3

Salmon GnRH

cDNA

CAG CAC TGG TCT TAT GGC TGG CTG CCT GGA

15

SEQ ID NO: 4

Salmon GnRH Analog

p-Glu-His-Trp-Ser-Tyr-D-Arg-Trp-Leu-Pro-aza-Gly-NH₂

20

SEQ ID NO: 5

Herring GnRH

cDNA

25

CAG CAC TGG TCT TAT GGC TGG CTG CCT GGA

SEQ ID NO: 6

Herring GnRH Analog

30 p-Glu-His-Trp-Ser-Tyr-D-Arg-Leu-Ser-Pro-aza-Gly-NH₂

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1	2	3	4	5	6	7	8	9	10	
p-Glu-	His-	Trp-	Ser-	Tyr-	Gly-	Leu-	Arg-	Pro-	Gly-NH ₂	Mammalian
p-Glu-	His-	Trp-	Ser-	Tyr-	Gly-	Leu-	Gln-	Pro-	Gly-NH ₂	Chicken I
p-Glu-	His-	Trp-	Ser-	Tyr-	Gly-	Trp-	Leu-	Pro-	Gly-NH ₂	Salmon
p-Glu-	His-	Trp-	Ser-	His-	Gly-	Trp-	Tyr-	Pro-	Gly-NH ₂	Chicken II
p-Glu-	His-	Trp-	Ser-	His-	Gly-	Trp-	Leu-	Pro-	Gly-NH ₂	Dogfish
p-Glu-	His-	Trp-	Ser-	His-	Gly-	Leu-	Ser-	Pro-	Gly-NH ₂	Herring
p-Glu-	His-	Trp-	Ser-	His-	Gly-	Leu-	Asn-	Pro-	Gly-NH ₂	Catfish
p-Glu-	His-	Trp-	Ser-	His-	Asp-	Trp-	Lys-	Pro-	Gly-NH ₂	Lamprey III
p-Glu-	His-	Tyr-	Ser-	Leu-	Glu-	Trp-	Lys-	Pro-	Gly-NH ₂	Lamprey I

Fig. 1

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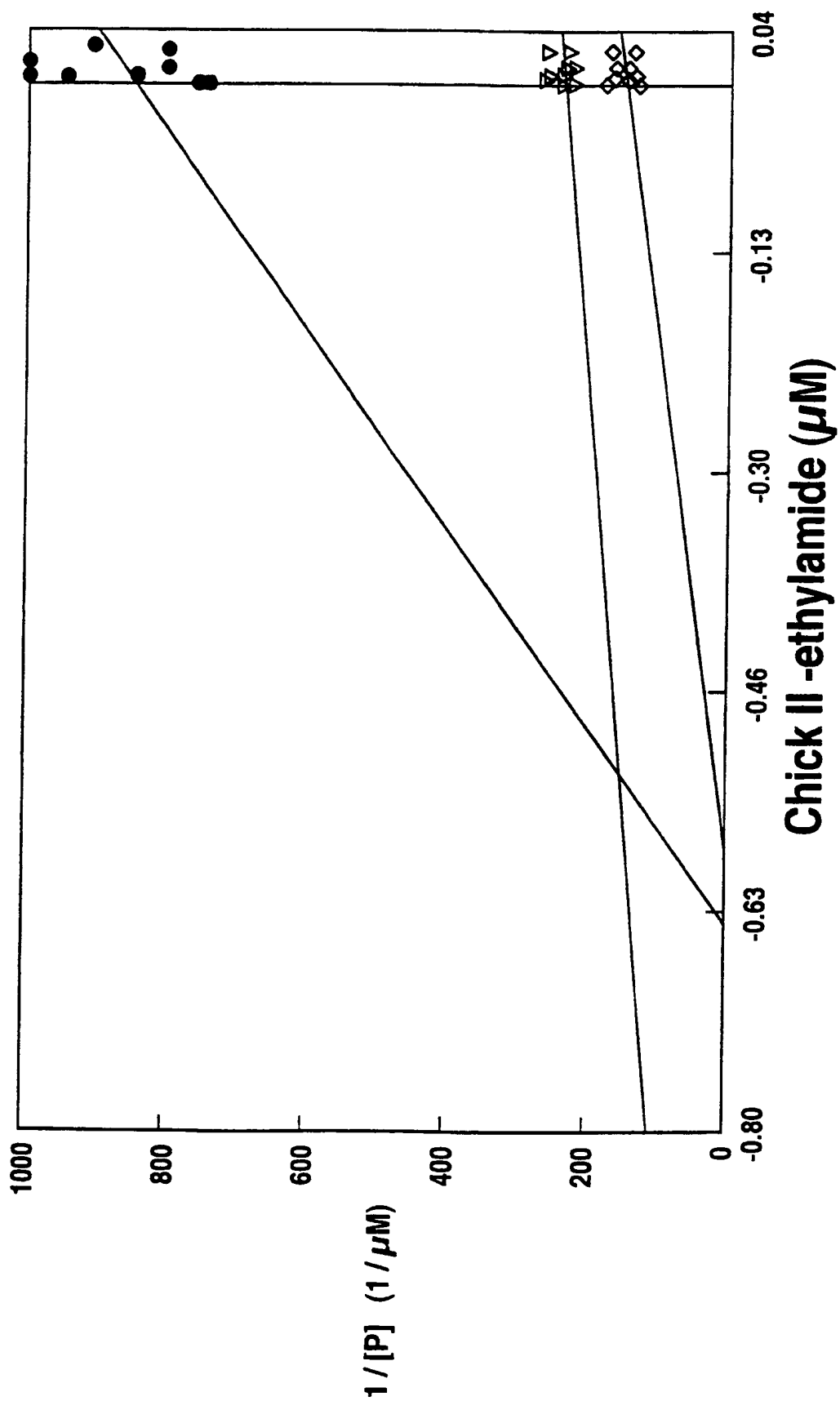


Fig. 2

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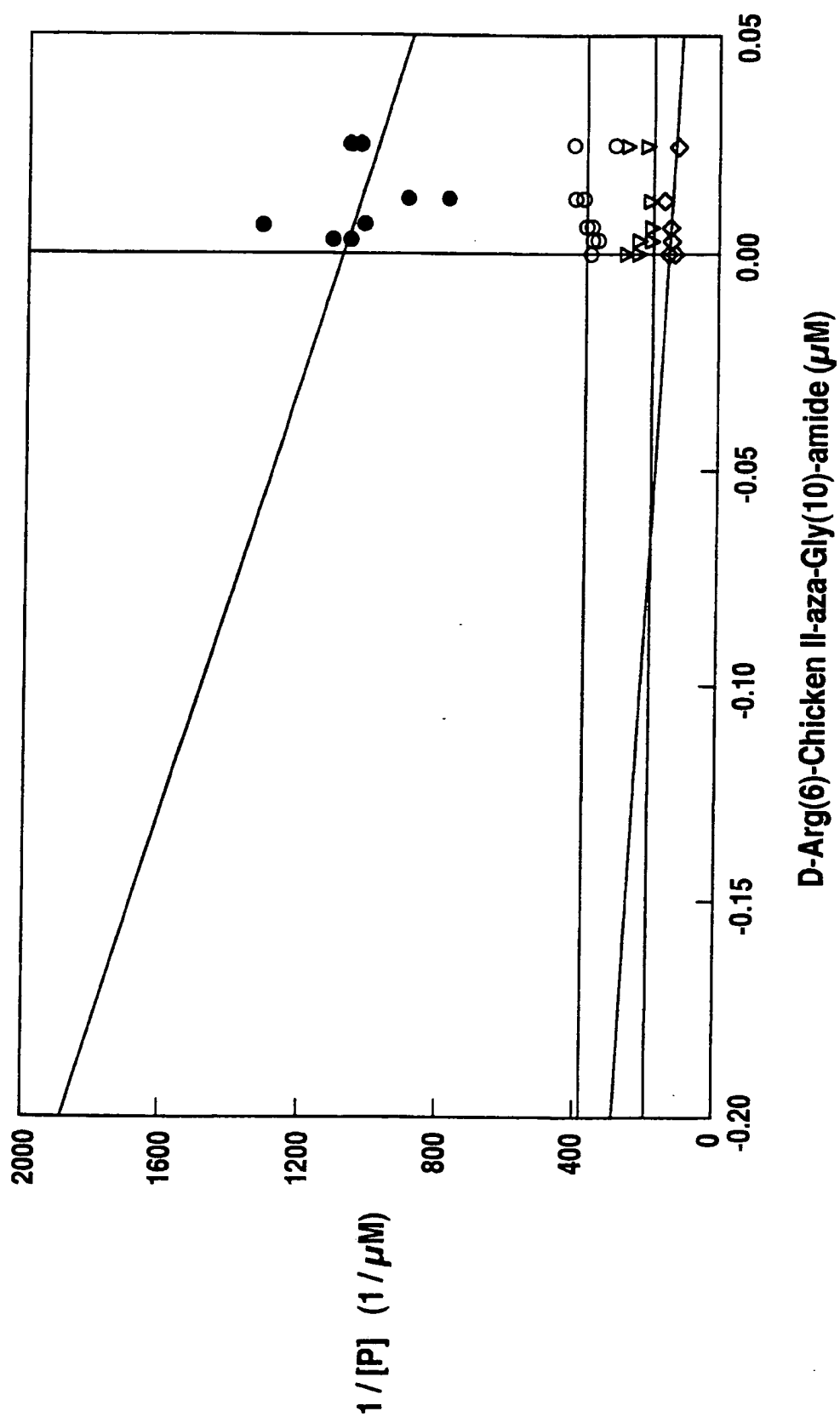


Fig. 3

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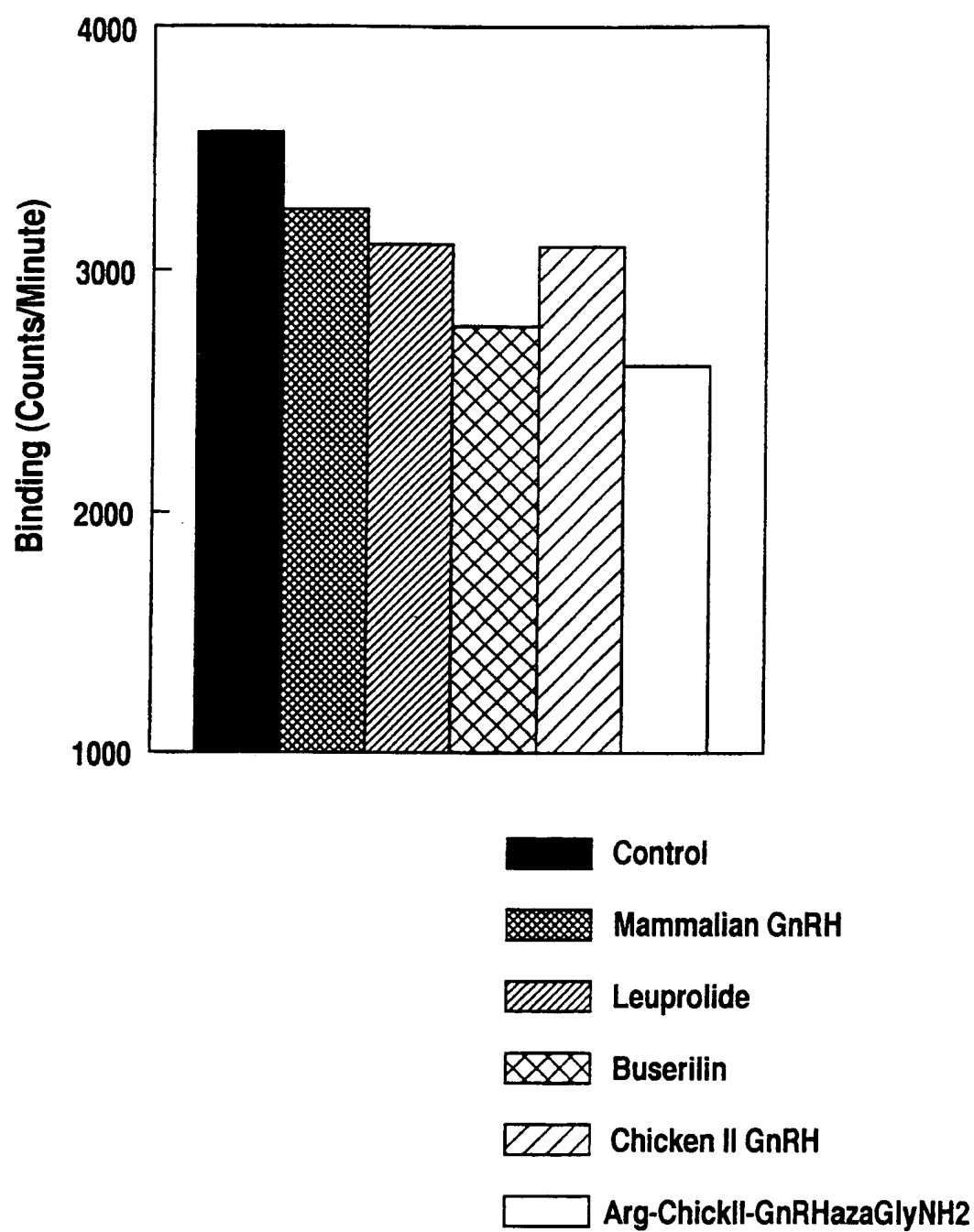


Fig. 4

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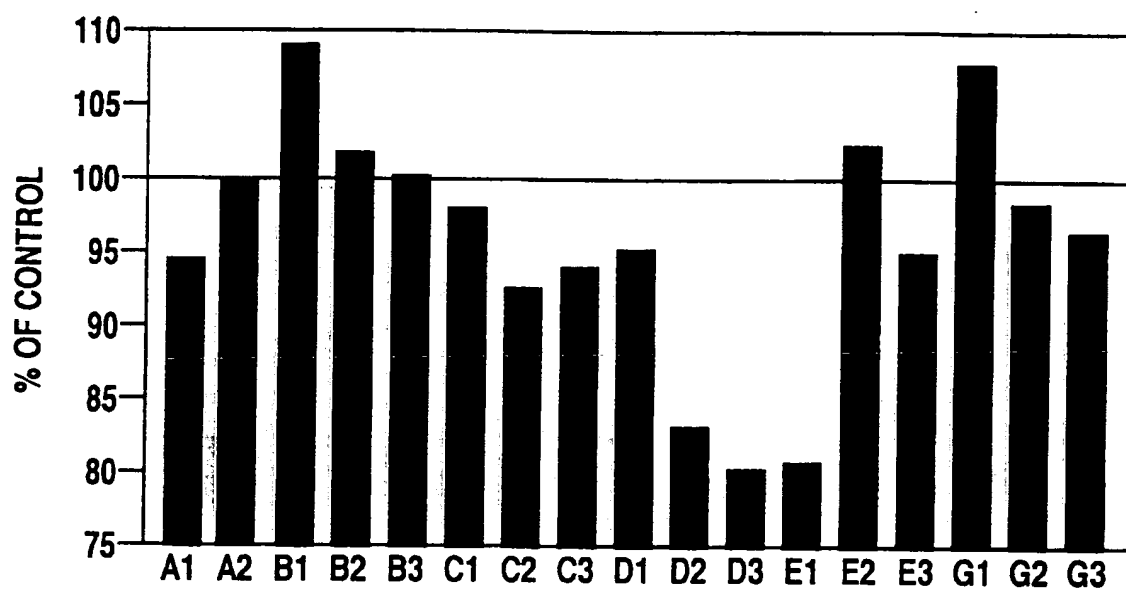


Fig. 5

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Analog of GnRH

Mammalian	30
Lamprey	20
Salmon	300
Chicken I	80
Chicken II	200
Chicken II EA (10)	130
Chicken II D-Arg (6), aza-Gly (10) amide	>200
Salmon II D-Arg (6), aza-Gly (10) amide	200
Mammalian D-Trp (6)	20
Mammalian EA (10)	70
Mammalian D-Trp (6), EA (10)	60
Mammalian D-Leu (6), EA (10)	80
Mammalian But-D-Ser (6), EA (10)	110
Mammalian Im-bzl-D-His (6), EA (10)	>200
Antide	120

Fig. 6

INTERNATIONAL SEARCH REPORT

International Application No
PCT/US 00/26575

A. CLASSIFICATION OF SUBJECT MATTER
IPC 7 A61K38/09 A61P35/00

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)
IPC 7 A61K C07K

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

EPO-Internal, BIOSIS, CHEM ABS Data, MEDLINE, WPI Data

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	NICHOLSON R I ET AL: "ANTITUMOR ACTIVITY OF FOLLIGEN A NOVEL GONADOTROPIN-RELEASING HORMONE ANALOGUE AGAINST DMBA-INDUCED TUMOURS IN THE RAT" TUMOR BIOLOGY, vol. 13, no. 1-2, 1992, pages 44-50, XP001002047 ISSN: 1010-4283	1-8, 17-19
Y	the whole document	9-16, 20, 21
Y	GB 2 237 571 A (MILLAR ROBERT PETER) 8 May 1991 (1991-05-08) abstract page 1 -page 3, paragraph 7 page 6, paragraph 2 - paragraph 3 --- -/--	9-16, 20, 21

☒ Further documents are listed in the continuation of box C.

☒ Patent family members are listed in annex.

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Date of the actual completion of the international search

18 May 2001

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29/05/2001

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INTERNATIONAL SEARCH REPORT

International Application No

PCT/US 00/26575

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	<p>KARTEN M J ET AL: "GONADOTROPIN-RELEASING HORMONE ANALOG DESIGN. STRUCTURE-FUNCTION STUDIES TOWARD THE DEVELOPMENT OF AGONISTS AND ANTAGONISTS: RATIONALE AND PERSPECTIVE"</p> <p>ENDOCRINE REVIEWS, US, BALTIMORE, MD, vol. 7, no. 1, 1986, pages 44-66, XP002038872</p> <p>the whole document</p> <p style="text-align: center;">----</p>	1-21
X	<p>US 5 760 000 A (HABIBI HAMID R)</p> <p>2 June 1998 (1998-06-02)</p> <p>abstract</p> <p>column 2, line 10 - line 24</p> <p>column 3, line 16 - line 25</p> <p>column 5, line 9 - line 52</p> <p>examples 2,8-10</p> <p style="text-align: center;">-----</p>	1-5, 17, 18

INTERNATIONAL SEARCH REPORT

Information on patent family members

International Application No

PCT/US 00/26575

Patent document cited in search report	Publication date	Patent family member(s)	Publication date
GB 2237571 A	08-05-1991	CA 2029018 A ZA 9008738 A	02-05-1991 28-08-1991
US 5760000 A	02-06-1998	AU 2265895 A WO 9531475 A	05-12-1995 23-11-1995